Call: 800-344-4100
Email: tech@lamotte.com

SAB/SAB

CODE 5551

USE

Selective isolation of fungi

APPLICATION

Sabouraud Dextrose Agar is used for cultivating pathogenic and commensal fungi and yeasts. The high dextrose concentration and acidic pH of the formula permits selectivity of fungi.

PADDLE AGARS

Note: Side 1 of each paddle is marked with an indented laser line.

Sabouraud Dextrose Agar (SAB) – (Color: Off-white) Sabouraud Dextrose Agar is used for cultivating pathogenic and commensal fungi and yeasts. The high dextrose concentration and acidic pH of the formula permits selectivity of fungi.

STORAGE / EXPIRATION

Store tightly sealed BioPaddles® in a cool, dry location. Shield from direct sunlight. Store BioPaddles® at room temperature (65 - 77°F/18 - 25°C). Avoid sudden temperature changes. Temperature fluctuations may result in condensation settling at the bottom of the vial. This will not affect the culture properties but could reduce the shelf-life or cause the agar to separate from the plastic paddle support. Do not refrigerate or store at temperatures above 80°F/27°C. Refrigeration may result in water condensation. Avoid freezing.

Refer to Best Before End date (See: BBE stamped on vial). Discard if paddle agar appears oxidized and darker than the expected color or if contaminants appear. The expiration date is based on medium in an intact container that is stored as directed.

AGAR VERIFICATION

These agars have been verified by EMSL Analytical, Inc. using E. coli and E. faecalis cultures. Documentation available upon request.

SAMPLING

LIQUID SAMPLING PROTOCOL

DIRECT IMMERSION PROTOCOL for Low Viscosity Liquids

1. Twist to remove paddle from vial. Do not touch agar surfaces.
2. Fill vial to 40 mL fill line with the liquid to be sampled and immerse paddle or immerse paddle directly in the sample. Both agar surfaces must be completely contacted. Allow at least 15 second contact time (30 seconds is optimal).
3. Remove paddle. Allow liquid to drain off both agar surfaces.
4. Replace paddle in vial.
5. Incubate.

SPREAD PROTOCOL for High Viscosity Liquids

1. Twist to remove paddle from vial. Do not touch agar surfaces.
2. Hold the contact agar surface on a horizontal plane. Deposit the liquid sample as a single drop approximately 1 cm from the handle boundary.
3. Position a sterile glass rod between the handle and the drop of sample. Bring the rod in contact with the drop to create a meniscus. Drag the rod over the agar surface toward the tip of the paddle.
4. Replace paddle in vial.
5. Incubate.

SURFACE SAMPLING PROTOCOL

Recovery Rate is about 50%

1. Twist paddle to remove from vial. Do not touch agar surfaces.
2. Touch the paddle surface (10 cm^2) to two different areas of the test surface to cover a total of 20 cm^2. Or touch the paddle to the surface once and multiply the colony count by 2.
3. Allow 15 second contact time.
4. Replace the paddle in the vial.
5. Incubate

AIR SAMPLING PROTOCOL

1. Twist to remove paddle from vial. Do not touch agar surfaces.
2. Invert paddle and insert the circular cap.
3. Expose for 15 minutes.
4. Replace paddle in vial.
5. Incubate.

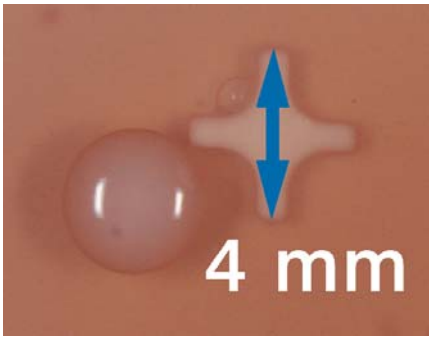
INCUBATION

Incubation of Paddle Growth	Incubation Temperature	Examine at:
Total Coliform / Bacteria	35 ± 2°C	24 to 48 hours
Total Coliform / Bacteria	Room Temperature	Up to 5 days
Yeast / Mold	25 to 30°C	48 hours up to 120 hours (5 days)
Yeast / Mold	Room Temperature	Up to 7 days

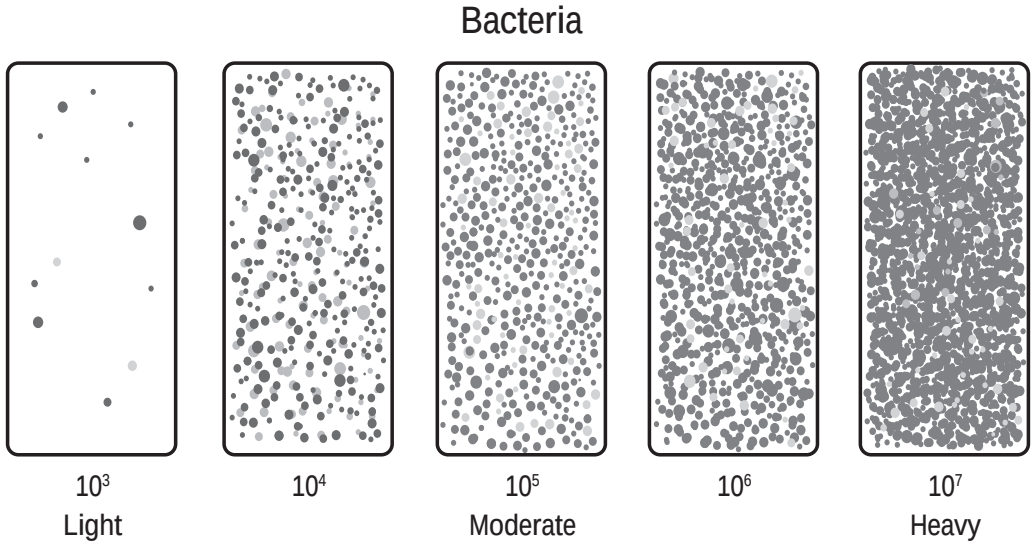
Note: Incubation of bacteria after 48 hours may produce confluent growth making enumeration more difficult.

COLONY MEASURING

Each BioPaddles® paddle has molded media attachment points that are 4 mm in length (point-to-point). This feature provides a useful guidepost to estimating nearby colony size.



ENUMERATION



Note: Estimation of lower counts is possible, but statistically difficult to justify. Use Light, Moderate and Heavy for Mold and Mildew growth. Mold and mildew colony growth is more confluent than bacterial growth and therefore more difficult to quantify. Use Light, Moderate, and Heavy for surface and air testing.

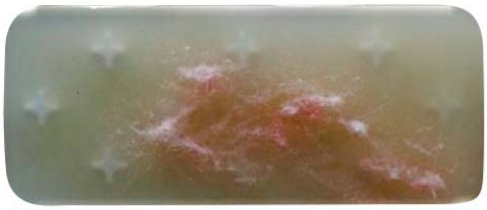
DISPOSAL

Twist to remove paddle from vial. Fill vial to 40 mL fill line with 1:9 dilution of household bleach (5.25% sodium hypochlorite). Replace paddle in vial. Allow 15 minute contact time. Remove paddle. Discard bleach solution. Replace paddle in vial and dispose. Alternatively, loosen cap and microwave for 30 seconds, autoclave, or incinerate.

IDENTIFICATION

An organism with Growth +++ will grow very well (non-fastidious) on the indicated media. An organism with Growth + is less likely to grow (fastidious), especially if crowded out by Growth +++ organisms. The media may not contain all of the nutrients that a Growth + organism needs in order to thrive.

Organism	Nutrient (NA) Agar
Aspergillus niger	 Growth: +++ Colony: Granular, white with yellow/gray hyphae, 2-5++ cm
Bacillus spp.	 Growth: +++ Colony: Translucent/off-white, smooth, spreading, 2-4 mm
Candida albicans	 Growth: +++ Colony: Cream, smooth, spreading, 2-6 mm

Cladosporium spp.	 Growth: + Colony: Granular/wooly, olive-brown, 2-5+ cm
Fusarium oxysporum	 Growth: +++ Colony: Wooly, initially white color changing to yellow, pink, red, or purple shades 4+ cm (confluent growth), fast growing
Muccor himaelis	 Growth: +++ Colony: Wooly, initially white, then white-yellow to various shades of gray to green showing lolipop sporangia, 3-9+ cm (confluent growth)
Penicillium chrysogenum (notatum)	 Growth: +++ Colony: Granular, velvet-like/powdery, flat, initially white then shades of green, blue-green, or yellow-green pigment, 3 - 9+ cm (confluent growth)

Penicillium roqueforti	 Growth: +++ Colony: Granular, velvet-like/powdery, flat, initially white, then blue or blue-green in color; white colony margins (growth areas), 2 - 5+ cm (confluent growth)
Rhizopus stolonifer	 Growth: +++ Colony: Cottony (dense growth), initially white, then becoming gray with black sporangia, 5+ cm (confluent growth), fast growing
Saccharomyces cerevisiae	 Growth: +++ Colony: Cream, circular, entire, raised to convex, 1-2 mm
Torula lactose	 Growth: +++ Colony: Transparent, circular to irregular, convex, dull, erose, 0.1 - 0.5 mm

GLOSSARY

Catalase Test	Catalase enzyme will react with hydrogen peroxide to produce oxygen if the bacteria is catalase positive.
Lactose Test	Lactose positive bacteria can ferment available lactose in the agar producing an acid which lowers the pH. Lactose negative bacteria are non-fermenting.
Indole Test	Biochemical test to determine the ability of an organism to split indole from the amino acid tryptophan. P. vulgaris is indole positive while P. mirabilis is indole negative.
Oxidase Test	Oxidase positive bacteria contain cytochrome c oxidase which will turn an indicator dark blue. In contact with oxidase negative bacteria, the indicator will remain colorless.
Urease Test	Bacteria containing urease will hydrolyze urea to ammonia and carbon dioxide causing an alkaline environment which changes the color of a pH indicator from yellow to fuchsia.
β-D-Glucoronidase Reaction	The presence of E. coli is determined when both β-D-Glucoronidase and Indole are positive, and the organism is gram negative.
Gram Staining	A method for differentiating bacteria into two groups – gram positive and gram negative – based on the chemical and physical properties of their cell walls. Often the first step in identifying bacteria.